# **WILEY** Online Proofing System Instructions

The Wiley Online Proofing System allows authors and proof reviewers to review PDF proofs, mark corrections, respond to queries, upload replacement figures, and submit these changes directly from the PDF proof from the locally saved file or while viewing it in your web browser.

- **1.** For the best experience reviewing your proof in the Wiley Online Proofing System please ensure you are connected to the internet. This will allow the PDF proof to connect to the central Wiley Online Proofing System server. If you are connected to the Wiley Online Proofing System server you should see the icon with a green check mark above in the yellow banner.
- Please review the article proof on the following pages and mark any corrections, changes, and query responses using the Annotation Tools outlined on the next 2 pages.
- **3.** To save your proof corrections, click the "Publish Comments" button appearing above in the yellow banner. Publishing your comments saves your corrections to the Wiley Online Proofing System server. Corrections don't have to be marked in one sitting, you can publish corrections and log back in at a later time to add more before you click the "Complete Proof Review" button below.
- **4.** If you need to supply additional or replacement files <u>bigger</u> than 5 Megabytes (MB) do not attach them directly to the PDF Proof, please click the "Upload Files" button to upload files:
- 5. When your proof review is complete and you are ready to submit corrections to the publisher, please click the "Complete Proof Review" button below:

**IMPORTANT:** <u>Do not click the "Complete Proof Review" button without replying to all author queries found on the last page of your proof</u>. Incomplete proof reviews will cause a delay in publication.

IMPORTANT: Once you click "Complete Proof Review" you will not be able to publish further corrections.

Connected Disconnected

• Ann	otatio	ns		
P	<b>B</b>	B		8-
T≈	Ŧ	푸	T	Ъ



#### USING e-ANNOTATION TOOLS FOR ELECTRONIC PROOF CORRECTION

# WILEY

Once you have Acrobat Reader open on your computer, click on the Comment tab at the right of the toolbar:



1. Replace (Ins) Tool – for replacing text.

Strikes a line through text and opens up a text box where replacement text can be entered.

#### How to use it

Ŧ

- Highlight a word or sentence.
- Click on the Replace (Ins) icon in the Annotations section.
- Type the replacement text into the blue box that appears.

# idard framework for the analysis of m

ic). Itereference		eneg
ble of strateg	<u> </u>	n f
aborofoomr	🗩 🕈 dthreshe	
inter of com	08/06/2011 15:58:17	O
: is that the st	, which led	of
nain compo	·	b
lanal and and		
level, are exc		
important w	OIRS ON CHUY BY	- ir
Mhencefort	h) <sup>1</sup> we open the 'h	lack h

3. Add note to text Tool – for highlighting a section to be changed to bold or italic.



Highlights text in yellow and opens up a text box where comments can be entered.

#### How to use it

- Highlight the relevant section of text.
- Click on the Add note to text icon in the Annotations section.
- Type instruction on what should be changed regarding the text into the yellow box that appears.

#### namic responses of mark ups ent with the **VAR** evidence

satior	♥ * dthreshe 08/06/2011 15:21:28	ith
y Ma		ell
and		ed
on n		ber
to a		on
stent	also with the demai	nd-

#### 2. Strikethrough (Del) Tool – for deleting text.



#### How to use it

- Highlight a word or sentence.
- Click on the Strikethrough (Del) icon in the Annotations section.

there is no room for extra profits and c ups are zero and the number of (et) values are not determined by Blanchard and Kiyotaki (1987), erfect competition in general equilibries ts of aggregate demand and supply classical framework assuming monop een an exogenous number of firms

4. Add sticky note Tool – for making notes at specific points in the text.
Marks a point in the proof where a comment needs to be highlighted.
How to use it
Click on the Add sticky note icon in the Annotations section.
Click at the point in the proof where the comment should be inserted.
Type the comment into the yellow box that appears.

тапи апи ѕиррту впоскъ. мозгот

alamir	<u> </u>	+
appunn	🖻 * dthreshe	U
numbe	08/06/2011 15:18:08	ff
dard fr	51	S
cy. Nev	)	(
ole of sti		N
ber of e	ompentors and the mi	р
is that t	he structure of the sect	0

#### USING e-ANNOTATION TOOLS FOR ELECTRONIC PROOF CORRECTION





# High intensity focused ultrasound as a potential new modality for the treatment of pigmentary skin disorder

S. Y. Choi<sup>1</sup>, K. H. Yoo<sup>2</sup>, C. T. Oh<sup>1,3</sup>, T. R. Kwon<sup>1,3</sup>, E. J. Choi<sup>3</sup>, J. Seok<sup>1</sup> and B. J. Kim<sup>1</sup>

<sup>1</sup>Department of Dermatology, Chung-Ang University College of Medicine, Seoul, South Korea,

<sup>2</sup>Department of Dermatology, Catholic Kwandong University International ST.Mary's Hospital, Incheon, South Korea and <sup>3</sup>Department of Medicine, Graduate school, Chung-Ang University, Seoul, South Korea

**Background/Purpose:** The clinical skin tightening benefits of high intensity focused ultrasound (HIFU) have been established, but its mechanism of action in pigmented skin disorders remains unknown. We macroscopically and histopathologically investigated dermatological changes after HIFU at different exposure doses in a UVB-induced guinea pig model of hyper-pigmentation.

**Methods:** We applied HIFU irradiation at 0.1 and 0.2 J/cm<sup>2</sup> to UVB-induced spotty hyperpigmentation in guinea pig skin. The therapeutic effects of HIFU were judged based on gross appearance using photography, dermoscopy, and chromametry during a period of 3 weeks after HIFU irradiation. Histological assessments were performed using Fontana-Masson staining 1 day before and 3 weeks after HIFU irradiation.

**Results:** Macroscopically, UVB-induced hyperpigmentation was significantly reduced 2 weeks after HIFU with 0.2 J/cm<sup>2</sup>,

C KIN COLOR is related to the amount and **D** distribution of melanin. Abnormal melanin accumulations on the unevenness of skin tone and the effects of surface imperfections have been discussed (1). Melanin accumulation can be due to many different causes such as hormonal imbalance or sun exposure, and may be either transient or permanent, as observed clinically in conditions such as melasma, chloasma, or lentigo. Unwanted pigmentation can cause patients to be uncomfortable, self-conscious, and reduce feelings of self-worth (2). Several treatments are used to reduce hyperpigmentation, including disruption of the distribution of melanosomes and inhibition of the tyrosinase enzyme.

High intensity focused ultrasound (HIFU) has been investigated as a tool for the treatment of solid benign and malignant tumors for many decades (3). Recently, HIFU was explored as a new treatment modality for skin tightening and and 3 weeks after HIFU with 0.1 J/cm<sup>2</sup>. Histopathologically, the heavy deposition of melanin in the epidermis induced by UVB exposure was reduced 3 weeks after HIFU irradiation.

**Conclusion:** We confirmed that HIFU has a positive effect on UVB-induced hyperpigmentation as well as mechanical destructive activity. We suggest that HIFU may be useful as an alternative modality for human patients suffering from skin pigmentary conditions.

**Key words:** high intense focused ultrasound – hyperpigmentation – pigmentation – UVB

© 2015 John Wiley & Sons A/S. Published by John Wiley & Sons Ltd Accepted for publication 17 May 2015

rejuvenation (4). High intensity focused ultrasound can produce small, micro-thermal lesions at precise depths in the dermis up to the fibromuscular layer, causing thermally induced contraction of collagen and tissue coagulation with subsequent collagenesis, while sparing the epidermis (5–7). To date, no experimental or clinical studies have evaluated the efficacy of HIFU for the treatment of pigmented skin lesions. In this study, we evaluated the effects of HIFU on hyperpigmentation using an animal model.

Guinea pigs are commonly used for studies of skin reactions to UV irradiation and of the protective effects of sunscreen on sunburn and tanning reactions in the skin. The effects of depigmenting agents on spotty pigmentation have also been evaluated using guinea pig models (8, 9). This study was undertaken to macroscopically and histopathologically investigate dermatological changes in UVB-induced guinea pig skin pigmentation after HIFU at different exposure doses. PE: Sudhakar S

CE: Prema

Dispatch: 27.5.15 No. of pages: 6

WILEY

12239 Manuscript No.

Journal Code

S R

# Materials and Methods

#### Guinea pig model

One 6-week-old female brownish guinea pig (Tokyo Laboratory Animals Science Co., Tokyo, Japan) was used in this study. The guinea pig was bred and housed under conventional conditions (temperature:  $23 \pm 3^{\circ}$ C, relative humidity: 55  $\pm$  15%) at the R&D Center of the College of Medicine at Chung-Ang University, Korea. All procedures were conducted in accordance with the guidelines of the Institutional Animal Care and Use Committee of Chung-Ang University (IRB number: 13-0020). After acclimatization for 7 days, the dorsal skin of the guinea pig was separated into four areas  $(2 \times 2 \text{ cm})$  as follows: area 1: no UVB-induced tanning (control), area 2: UVB-induced tanning (control), area 3: UVB-induced tanning with HIFU at 0.1J/cm<sup>2</sup>, and area 4: UVB-induced tanning with HIFU at 0.2 J/cm<sup>2</sup>.

## UVB irradiation regimen

The guinea pig was anesthetized with Zoletil 50 (Virbac S.A, France) (40 mg/kg) and Rompun (Bayer, Korea) (5 mg/kg) in saline (Huons, Korea). To develop pigmentation, the back of the guinea pig was cleanly shaved with electric clippers. The guinea pig was exposed to weekly sessions of narrow band UVB (NB-UVB) irradiation for 4 weeks at a dose of 490 mJ/cm<sup>2</sup> per session using a NB-UVB lamp (Dermalight<sup>®</sup>80, National Biological Corp., OH, USA).

## HIFU ultrasound device protocol

A HIFU device (Ultraformer2<sup>®</sup>, Classys Inc., Seoul, Korea) was used in this study. An ultrasound probe was connected to a generator system operating in the MHz frequency regime. The ultrasound energy was coupled from the transducer (operating at 7 MHz) to skin by ultrasound coupling gel applied to the skin surface. The nominal focal depth for this study was 1.5 mm below the skin surface. Each probe delivered a set of pulses in a linear array, pulses spaced 1.0-2.0 mm apart, and an entire linear array was up to 25 mm long. The spacing of pulses within each linear array was set at 1 mm, resulting in 25 thermal coagulative zones created with each probe discharge. Linear arrays were spaced in parallel at 1-mm intervals. Ultrasound transmission gel (Supersonic<sup>®</sup>, Sungheung Co., Korea) was applied to the skin, and handpiece **2** was pressed perpendicularly, uniformly and firmly to the skin surface. The guinea pig was treated only once with a 7-MHz, 1.5-mm handpiece, at 0.1J/cm<sup>2</sup> and 0.2J/cm<sup>2</sup>. After treatment, the ultrasound transmission gel was wiped off of the guinea pig's skin. The treated skin showed mild redness and swelling that persisted for several days.

# Evaluation of tanning reduction

We evaluated tanning reduction 1, 2, and 3 weeks after HIFU treatment using photography, dermoscopy, and chromametry. Clinical changes were measured using digital photographs (Canon 3000D, Canon Inc., Tokyo, Japan). We used a dermoscope to produce images with enhanced magnification (DermLite Pro, CA, USA). The lightening effect was determined by measuring the L\* value with a CR-10 reflectance spectrophotometer (Konica Minolta Sensing, Inc., Sakai, Osaka, Japan) as a chromameter. The L\* value (luminance) defines the relative lightness ranging from total black  $(L^* = 0)$  to total white  $(L^* = 100)$ . The blanching effect was quantified by the increase in L\* value:  $\Delta L^* = L^*$  (on the measuring day) – L\* (on the first day of the test, before HIFU treatment).

## Histological analysis

Three weeks after HIFU treatment, the guinea pig was sacrificed and skin samples were removed from each quadrant of the test site. Samples were fixed in 10% formaldehyde, embedded in paraffin, and stained with standard hematoxylin and eosin (H&E). Changes in melanin deposition were measured by Fontana-Masson (FM) staining. All staining was examined under a phase-contrast microscope (Eclipse TS100<sup>®</sup>, Nikon Instruments Inc., Melville, NY, USA).

## Statistical analysis

Data are presented as mean  $\pm$  standard deviation. Statistical comparisons between the treated and untreated areas were performed using one-way ANOVA followed by Tukey's post hoc test for direct comparisons between groups. *P* values < 0.05 and <0.01 were considered statistically significant.

# Results

#### Clinical and dermoscopic changes

In both digital photographs and dermoscopic pictures, UVB-induced hyperpigmentation started to decrease 1 week after HIFU treatment in areas 3 and 4, while no reduction occurred in area 2. At 3 weeks after HIFU treatment, the tanning induced by UVB radiation was markedly reduced in areas 3 and 4. Compared with area 3, tanning in area 4 decreased more quickly (Figs 1 and 2).

#### Changes of brightness index

Compared with  $\Delta L^*$  in area 2, the L\* values of areas 3 and 4 were significantly decreased from baseline (before HIFU treatment), at 3 weeks and 2 weeks after HIFU treatment, respectively (P < 0.01) (Fig. 3). The details of the L\* values are shown in Table 1.

#### High intensity ultrasound for dyspigmentation

#### Histological changes

Microscopic examinations of H&E-stained sections confirmed that there were no signs of inflammatory or necrotic reactions in any of the four tested areas (Fig. 4). In FM-stained sections, marked increases of melanin in the basal layer of the epidermis were detected in area 2. However, in areas 3 and 4, heavy deposition of melanin in the epidermis induced by UVB exposure was reduced compared with area 2 (Fig. 5).

#### Discussion

Melanin pigment is a heterogeneous biopolymer synthesized from intermediate products derived from dopaquinone in the epidermis. The perceived color of skin is determined by the ratio of eumelanins to phaeomelanins, and in part by blood within the dermis. Exposure of skin to UVB irradiation upregulates the synthesis of melanocyte tyrosinase, regulating in increased melanogenesis and, thus tanning (10, 11).

In the present study, depigmentation with HIFU was investigated macroscopically and his-



Fig. 1. Digital photographs show that tanning induced by UVB exposure was markedly reduced in areas 3 and 4 at 3 weeks after HIFU treat-



Fig. 2. Dermoscopic pictures show that tanning induced by UVB exposure was markedly reduced in areas 3 and 4 at 3 weeks after HIFU treat- **5** ment.



Fig. 3. The L\* values of areas three and four were significantly decreased from baseline at 3 weeks and 2 weeks after HIFU treatment, respectively, compared with area 2 ( $P < 0.01^{**}$ ).

topathologically, using a UVB-induced hyperpigmentation model in the skin of a guinea pig. We macroscopically confirmed that UVBinduced hyperpigmentation significantly decreased after HIFU treatment with 0.1 J/cm<sup>2</sup> and 0.2 J/cm<sup>2</sup>. The effects of HIFU on UVBinduced hyperpigmentation were enhanced when applied with 0.2 J/cm<sup>2</sup> energy compared with 0.1 J/cm<sup>2</sup> energy. Histologically, we also confirmed that the melanin deposition in the epidermis induced by UVB exposure was markedly reduced after HIFU treatment with 0.1 J/

TABLE 1. The details of the L\* values

Days		0	7	14	21
Group 1	L*1	66.7	65.4	66.3	66.2
	L*2	66.4	65.5	65.4	65
	L*3	65.6	65.9	65.4	65.6
	L* Mean	66.23	65.6	65.7	65.6
	L* SD	0.57	0.26	0.52	0.6
Group 2	L*1	56	51.3	55.9	54.9
	L*2	57.5	54.4	55.3	55.6
	L*3	56.7	54.4	55.1	55.3
	L* Mean	56.73	53.36	55.43	55.27
	L* SD	0.75	1.79	0.42	0.35
Group 3	L*1	55	55	55.7	57.2
	L*2	54.7	54.8	56.4	56.8
	L*3	55.1	53.9	56.2	57.4
	L* Mean	54.93	54.56	56.1	57.13
	L* SD	0.21	0.56	0.36	0.31
Group 4	L*1	52.3	55.7	58	59
	L*2	54.6	58	57.2	57.7
	L*3	53.6	56.4	58.2	58.8
	L* Mean	53.5	56.7	57.8	58.5
	L* SD	1.15	1.18	0.53	0.7

SD, Standard deviation

cm<sup>2</sup> and 0.2 J/cm<sup>2</sup>. Therefore, we suggest that HIFU has skin lightening effects on areas with UVB-induced hyperpigmentation.

HIFU has recently been used for skin tightening and rejuvenation. Usually, 3- and 4.5-mm transducers are applied to deliver energy to the deep



Fig. 4. There were no signs of inflammatory or necrotic reactions of skin tissue (hematoxylin & eosin stain).



Fig. 5. Heavy deposition of melanin in the epidermis induced by UVB exposure was reduced in areas 3 and 4 (Fontana-Masson stain).

7

6

dermis, subcutis, and fibromuscular layer. Epidermal injury is minimized and ultrasound energy is directed into the deep skin tissue, resulting in well-defined thermal injury zones (5–7).

The mechanism underlying the lightening effects of HIFU is not understood. We hypothesize that when using a HIFU 1.5-mm transducer, the ultrasound energy is delivered beneath the dermoepidermal junction and upper dermis. The ultrasound waves induce vibrations in the composite molecules within skin tissue during propagation, and the friction that develops between intrinsic molecules is the source of the generated heat (12). We then propose that the mechanical destructive effects induced by vibration and friction are what eliminate melanin and pigmented debris from the epidermis and upper dermis.

Similarly, melasma has been successfully treated with fractional resurfacing lasers. Fractional photothermolysis may induce ultrastructural changes, resulting in decreases in the numbers of melanocytes and melanin granules within keratinocytes (13, 14). HIFU is similar to fractional laser resurfacing in that thermal lesions are created, but is unique in that the thermal lesions are created below the surface and can be of variable geometry (10). As fractional resurfacing lasers have been used for the treatment of pigmented lesions including melasma, HIFU may be effective due to similar mechanisms for the elimination of melanocytes and melanin, and may be helpful to treat skin pigmentary conditions.

Our results demonstrate that a single session of HIFU treatment using a 1.5-mm-depth transducer is effective for improving UVB-induced hyperpigmentation in an animal model. The major limitation of this study is the use of tanning loss to assess depigmentation capacity. This

# method is widely used for testing pigmentary skin problems, but does not measure the ability to reduce long-lasting pigmentation such as freckles or melasma. Based on this animal study, we suggest that HIFU may be useful as an alternative modality for the treatment of skin pigmentary conditions in human patients. Further clinical studies are necessary to evaluate the effects of HIFU on pigmentary skin disorders.

# Acknowledgements

This study was supported by the Infrastructure Program for New-growth Industries (10044186, Development of Smart Beauty Devices Technology and Establishment of Commercialization Support Center), and funded by the Ministry of Trade, Industry & Energy (MI, Korea).

*Funding sources:* None. *Conflicts of interest:* None

# References

- 1. Calzavara-Pinton PG, Ortel B. Pigmentation after solar radiation. In: Giacomoni PU, ed. Biophysical and Physiological Effects of Solar Radiation on Human Skin, Cambridge: The Royal Society of Chemistry; 2007: 65–97.
- 2. Pawaskar MD, Parikh P, Markowski T et al. Melasma and its impact on health-related quality of life in Hispanic women. J Dermatol Treat 2007; 18: 5–9.
- Laubach HJ, Makin IR, Barthe PG, Slayton MH, Manstein D. Intense focused ultrasound: evaluation of a new treatment modality for precise microcoagulation within the skin. Deramtol Surg 2008; 34: 727–734.
- 4. Lee HJ, Lee KR, Park JY, Yoon MS, Lee SE. The efficacy and safety of intense focused ultrasound in the treatment of enlarged facial pores in Asian skin. J Dermatolog Treat 2015; 26: 73–77.
- Alam M, White LE, Marin N, Witherspoon J, Yoo S, West DP. Ultrasound tightening of facial and neck skin: a rater-blinded prospective cohort study. J Am Acad Dermatol 2010; 62: 262–269.

- 6. Suh DH, Shin MK, Lee SJ, Rho JH, Lee MH, Kim NI, Song KY. Intense focused ultrasound tightening in Asian skin: clinical and pathologic results. Dermatol Surg 2011; 37: 1595–1602.
- 7. Lee HS, Jang WS, Cha YJ, Choi YG, Tak Y, Hwang E, Kim BJ, Kim MN. Multiple pass ultrasound tightening of skin laxity of the lower face and neck. Dermatol Surg 2012; 38: 20–27.
- 8. Anbar TS, El-Ammawi TS, Barakat MT, Abdel-Rahman AT, Fawzy A. A new morphometric technique for assessment of melanization in skin of guinea pigs. Photodermatol Photoimmunol Photomed 2012; 28: 42–46.
- 9. Anbar TS, El-Ammawi TS, Barakat M, Fawzy A. Skin pigmentation after NB-UVB and three analogues of prostaglandin F(2alpha) in guinea pigs: a comparative study. J Eur Acad Dermatol Venereol 2010; 24: 28–31.
- Prota G. Melanins and Melanogenesis. ????: New York Academic Press 1992.
- 11. Ito S, Wakamatsu K, Ozeki H. Chemical analysis of melanins and

its application to the study of the regulation of melanogenesis. Pigement Cell Res 2000; 13: 103–109.

- 12. Whte WM, Makin IR, Slayton MH, Barthe PG, Gliklich R. Selective transcutaneous delivery of energy to porcine soft tissues using Intense Ultrasound (IUS). Lasers Surg Med 2008; 40: 67–75.
- Tannous ZS, Astner S. Utilizing fractional resurfacing in the treatment of therapy-resistant melasma. J Cosmet Laser Ther 2005; 7: 39–43.
- Goldberg DJ, Berlin AL, Phelps R. Histologic and ultrastructural analysis of melasma after fractional resurfacing. Lasers Surg Med 2008; 40: 134–138.

Address:

B. J. Kim, MD Dept. of Dermatology, Chung Ang University Hospital 224-1 Heukseok-dong, Dongjakku, Seoul 156-755 South Korea Tel: +82-2-6299-1525 Fax: +82-2-823-1049 e-mail: beomjoon@unitel.co.kr

# **Author Query Form**

# Journal: SRT Article: 12239

#### Dear Author,

During the copy-editing of your paper, the following queries arose. Please respond to these by marking up your proofs with the necessary changes/additions. Please write your answers on the query sheet if there is insufficient space on the page proofs. Please write clearly and follow the conventions shown on the attached corrections sheet. If returning the proof by fax do not write too close to the paper's edge. Please remember that illegible mark-ups may delay publication.

Many thanks for your assistance.

Query reference	Query	Remarks
1	AUTHOR: Please confirm that given names (red) and surnames/family names (green) have been identified correctly.	
2	AUTHOR: Please give address information for Sungheung Co.: city.	
3	AUTHOR: Please provide the publisher location for reference [10].	
4	AUTHOR: Figure 1 has been saved at a low resolution of 185 dpi. Please resupply at 300 dpi. Check required artwork specifications at http://authorservices.wiley.com/bauthor/ illustration.asp	
5	AUTHOR: Figure 2 has been saved at a low resolution of 185 dpi. Please resupply at 300 dpi. Check required artwork specifications at http://authorservices.wiley.com/bauthor/ illustration.asp	
6	AUTHOR: Figure 4 has been saved at a low resolution of 198 dpi. Please resupply at 600 dpi. Check required artwork specifications at http://authorservices.wiley.com/bauthor/ illustration.asp	
7	AUTHOR: Figure 5 has been saved at a low resolution of 189 dpi. Please resupply at 300 dpi. Check required artwork specifications at http://authorservices.wiley.com/bauthor/ illustration.asp	