

ORIGINAL ARTICLE

Asciminib in Chronic Myeloid Leukemia after ABL Kinase Inhibitor Failure

T.P. Hughes, M.J. Mauro, J.E. Cortes, H. Minami, D. Rea, D.J. DeAngelo, M. Breccia, Y.-T. Goh, M. Talpaz, A. Hochhaus, P. le Coutre, O. Ottmann, M.C. Heinrich, J.L. Steegmann, M.W.N. Deininger, J.J.W.M. Janssen, F.-X. Mahon, Y. Minami, D. Yeung, D.M. Ross, M.S. Tallman, J.H. Park, B.J. Druker, D. Hynds, Y. Duan, C. Meille, F. Hourcade-Potelleret, K.G. Vanasse, F. Lang, and D.-W. Kim

ABSTRACT

BACKGROUND

Asciminib is an allosteric inhibitor that binds a myristoyl site of the BCR-ABL1 protein, locking BCR-ABL1 into an inactive conformation through a mechanism distinct from those for all other ABL kinase inhibitors. Asciminib targets both native and mutated BCR-ABL1, including the gatekeeper T315I mutant. The safety and antileukemic activity of asciminib in patients with Philadelphia chromosome-positive leukemia are unknown.

METHODS

In this phase 1, dose-escalation study, we enrolled 141 patients with chronic-phase and 9 with accelerated-phase chronic myeloid leukemia (CML) who had resistance to or unacceptable side effects from at least two previous ATP-competitive tyrosine kinase inhibitors (TKIs). The primary objective was to determine the maximum tolerated dose or the recommended dose (or both) of asciminib. Asciminib was administered once or twice daily (at doses of 10 to 200 mg). The median follow-up was 14 months.

RESULTS

Patients were heavily pretreated; 70% (105 of 150 patients) had received at least three TKIs. The maximum tolerated dose of asciminib was not reached. Among patients with chronic-phase CML, 34 (92%) with a hematologic relapse had a complete hematologic response; 31 (54%) without a complete cytogenetic response at baseline had a complete cytogenetic response. A major molecular response was achieved or maintained by 12 months in 48% of patients who could be evaluated, including 8 of 14 (57%) deemed to have resistance to or unacceptable side effects from ponatinib. A major molecular response was achieved or maintained by 12 months in 5 patients (28%) with a T315I mutation at baseline. Clinical responses were durable; a major molecular response was maintained in 40 of 44 patients. Dose-limiting toxic effects included asymptomatic elevations in the lipase level and clinical pancreatitis. Common adverse events included fatigue, headache, arthralgia, hypertension, and thrombocytopenia.

CONCLUSIONS

Asciminib was active in heavily pretreated patients with CML who had resistance to or unacceptable side effects from TKIs, including patients in whom ponatinib had failed and those with a T315I mutation. (Funded by Novartis Pharmaceuticals; ClinicalTrials.gov number, NCT02081378.)

The authors' full names, academic degrees, and affiliations are listed in the Appendix. Address reprint requests to Dr. Hughes at the South Australian Health and Medical Research Institute, North Terrace, Adelaide, SA, Australia, or at tim.hughes@sahmri.com.

Drs. Hughes and Mauro and Drs. Lang and Kim contributed equally to this article.

N Engl J Med 2019;381:2315-26.

DOI: 10.1056/NEJMoa1902328

Copyright © 2019 Massachusetts Medical Society.

FAILURE OF TYROSINE KINASE INHIBITOR (TKI) therapy in patients with Philadelphia chromosome (Ph)-positive chronic myeloid leukemia (CML) may result from resistance to or unacceptable side effects from the drug or both. Currently approved TKIs mainly target the ATP-binding site of BCR-ABL1, and approximately half of clinical resistance is associated with the acquisition of mutations in this region of the kinase, resulting in conformational changes that render TKIs inactive.¹⁻⁶ The “gatekeeper” T315I mutation, reported in approximately 20% of patients with mutations, is of particular concern because it is associated with resistance to all clinically available TKIs except ponatinib.⁷⁻⁹ Unacceptable side effects from TKIs also occur in approximately 25% of patients, with increasing recognition that patients receiving second- and third-generation TKIs are at risk for vascular and pulmonary toxic effects.¹⁰⁻¹⁴

Asciminib (ABL001) is a potent, specific, orally bioavailable BCR-ABL1 inhibitor that is distinct from approved ABL1 kinase inhibitors in that it does not bind to the ATP-binding site of the kinase. In contrast, asciminib acts as an allosteric inhibitor and engages a vacant pocket at a site of the kinase domain normally occupied by the myristoylated N-terminal of ABL1 — a motif that serves as an allosteric negative regulatory element lost on fusion of ABL1 to BCR (Fig. 1).

By binding the myristoyl site, asciminib mimics myristate and restores inhibition of kinase activity. Owing to the distinct conformation of the myristoyl pocket, asciminib has high selectivity for only ABL1 and, hypothetically, ABL2 kinases, with low-nanomolar-range activity against unmutated BCR-ABL1 and all clinically observed ATP-site mutants, including T315I.^{15,16} We hypothesized that asciminib may produce clinically significant responses in patients with CML in whom multiple approved TKIs have failed.

We conducted a phase 1 study to determine the safety, maximum tolerated dose or recommended dose, pharmacokinetics, and antileukemic activity of asciminib in patients with Ph-positive leukemia after failure of multiple approved TKIs. Here we report on patients with CML in the chronic or accelerated phase.

METHODS

STUDY OVERSIGHT

The study was designed collaboratively by the sponsor (Novartis Pharmaceuticals) and study investigators. The sponsor collected the data and analyzed them in conjunction with the authors. The first two authors wrote the first draft of the manuscript. Editorial support was provided by ArticulateScience and funded by the sponsor. All authors vouch for the accuracy and completeness

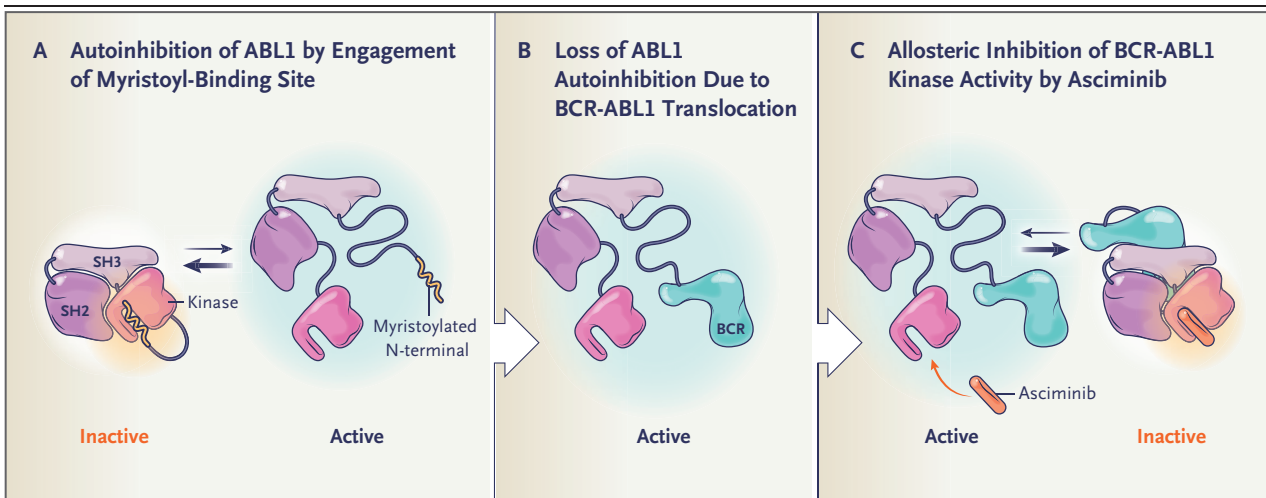


Figure 1. Binding of the Myristoyl Site of the BCR-ABL1 Protein by Asciminib.

Autoinhibition of the ABL1 kinase occurs through engagement of the myristoyl-binding site by the myristoylated N-terminal — a negative regulatory motif that locks the ABL1 kinase in the inactive state (Panel A). On fusion of ABL1 to BCR, the myristoylated N-terminal is lost and the ABL1 kinase is activated (Panel B). By allosterically binding the myristoyl site, asciminib mimics myristate and restores inhibition of BCR-ABL1 kinase activity (Panel C).

of the data and for the fidelity of the study to the protocol (available with the full text of this article at NEJM.org).

PATIENTS

Patients were eligible if they were 18 years of age or older, had Ph-positive chronic-phase or accelerated-phase CML, and had hematologic, cytogenetic, or molecular disease that was relapsed or refractory to at least two different TKIs before study entry or had unacceptable side effects from the TKIs, as determined by investigators according to standard criteria.¹⁷ Patients with a *BCR-ABL1* T315I mutation were eligible after they had received at least one TKI if no other effective therapy was available. Additional cohorts of patients were subsequently enrolled through a protocol amendment (Fig. S1 in the Supplementary Appendix, available at NEJM.org).

STUDY DESIGN

The primary objective was to determine the maximum tolerated dose or the recommended dose (or both) of asciminib administered twice daily in patients with chronic-phase or accelerated-phase CML. Secondary objectives included assessments of safety, pharmacokinetics, and efficacy. The study included a dose-escalation phase and an expansion phase for patients treated at either the maximum tolerated dose or the recommended dose.

PHARMACOKINETICS

Data on pharmacokinetics are presented for 110 patients from cycle 1, day 1 onward (Table S1). A two-compartment pharmacokinetic–pharmacodynamic model with linear elimination and distribution was used to inform the recommended dose selection (see the Supplementary Appendix).

ANTILEUKEMIC ACTIVITY

Complete blood counts were performed regularly to assess hematologic response. Bone marrow aspirations for morphologic and cytogenetic analyses were performed before therapy and during the study in all patients with *BCR-ABL1* transcript levels of more than 1% on the International Scale (IS) or loss of a complete hematologic response, every 3 months in patients without a complete cytogenetic response, and as clinically indicated. Real-time quantitative reverse-transcriptase–polymerase-chain-reaction assays

for molecular response were performed every 3 months and as clinically indicated.

Responses were defined according to standard criteria.^{18–22} Molecular responses were assessed with the ratio of *BCR-ABL1* to *ABL1* measured on the International Scale (*BCR-ABL1*^{IS}).²² Molecular response was calculated for patients with typical b2a2 or b3a2 *BCR-ABL1* transcripts only. Changes in molecular-response category from baseline were assessed with the use of intervals of 1-log changes in *BCR-ABL1* transcript levels by 6 or 12 months (see the Supplementary Appendix). *BCR-ABL1* myristoyl-pocket mutations were assessed by means of bidirectional Sanger and next-generation sequencing in patients who could be evaluated and who had molecular disease progression or loss of a complete cytogenetic response at any time and in patients who had received asciminib for at least 12 months at the time of analysis (Fig. S2). Molecular assessments were performed centrally (MolecularMD, Portland, Oregon).

STATISTICAL ANALYSIS

A Bayesian logistic-regression model^{23,24} was used to estimate the posterior distribution of probabilities of dose-limiting toxic effects at various doses after each patient cohort in dose escalation and to determine the maximum tolerated dose (Fig. S3).

RESULTS

PATIENTS

Treatment status and patient characteristics are shown in Table 1 and Table S2. From May 2014 through September 2017, a total of 141 patients with chronic-phase CML and 9 with accelerated-phase CML were treated with asciminib monotherapy, with a median follow-up of 59 weeks (range, 0.1 to 167) at the time of analysis. Of all 150 patients enrolled, 105 (70%) had received at least three previous TKIs. At study entry, 46 patients (31%) had at least one *BCR-ABL1* kinase domain mutation, the most frequent being T315I (in 33 patients [22%]). As of September 1, 2017, a total of 110 patients (73%) were continuing study treatment. Of patients who discontinued treatment, 1 died (after a blast crisis developed).

SAFETY PROFILE

In patients with CML treated on a twice-daily schedule, seven dose levels were investigated:

Table 1. Treatment Status and Demographic and Clinical Characteristics of the Patients at Baseline, According to Asciminib Dosing Schedule.*

Variable	Chronic-Phase CML				Accelerated-Phase CML			
	No T3151 Mutation		T3151 Mutation		No T3151 Mutation		T3151 Mutation	
	2x/Day (N=68)	1x/Day (N=45)	Combined 1x/Day and 2x/Day (N=113)	200 mg 2x/Day (N=9)	Combined 1x/Day and 2x/Day (N=28)	Combined 1x/Day and 2x/Day (N=4)	Combined 1x/Day and 2x/Day (N=5)	
Continued to receive asciminib at time of analysis	48 (71)	40 (89)	88 (78)	9 (100)	19 (68)	2 (50)	1 (20)	
Discontinued asciminib	20 (29)	5 (11)	25 (22)	0	9 (32)	2 (50)	4 (80)	
Adverse event	6 (9)	1 (2)	7 (6)	0	3 (11)	0	0	
Death	1 (1)	0	1 (1)	0	0	0	0	
Physician decision	4 (6)	2 (4)	6 (5)	0	1 (4)	0	1 (20)	
Progressive disease	7 (10)	2 (4)	9 (8)	0	3 (11)	2 (50)	3 (60)	
Patient or guardian decision	2 (3)	0	2 (2)	0	2 (7)	0	0	
ECOG performance-status score†								
0	52 (76)	30 (67)	82 (73)	5 (56)	21 (75)	3 (75)	4 (80)	
1	15 (22)	15 (33)	30 (27)	4 (44)	6 (21)	1 (25)	1 (20)	
2	1 (1)	0	1 (1)	0	1 (4)	0	0	
No. of previous TKIs								
1	2 (3)	0	2 (2)	0	4 (14)	0	0	
2	20 (29)	10 (22)	30 (27)	2 (22)	8 (29)	1 (25)	0	
≥3	46 (68)	35 (78)	81 (72)	7 (78)	16 (57)	3 (75)	5 (100)	
Previous TKI								
Imatinib	46 (68)	37 (82)	83 (73)	7 (78)	21 (75)	4 (100)	4 (80)	
Nilotinib	49 (72)	37 (82)	86 (76)	7 (78)	15 (54)	3 (75)	5 (100)	
Dasatinib	57 (84)	41 (91)	98 (87)	8 (89)	19 (68)	4 (100)	5 (100)	
Bosutinib	23 (34)	20 (44)	43 (38)	1 (11)	5 (18)	1 (25)	1 (20)	
Radotinib	5 (7)	1 (2)	6 (5)	2 (22)	4 (14)	0	0	
Ponatinib	20 (29)	14 (31)	34 (30)	7 (78)	15 (54)	1 (25)	2 (40)	
BCR-ABL1 transcript								
Typical	63 (93)	42 (93)	105 (93)	9 (100)	27 (96)	4 (100)	5 (100)	
Atypical	3 (4)	3 (7)	6 (5)	0	1 (4)	0	0	
Unknown	2 (3)	0	2 (2)	0	0	0	0	

* Percentages are based on the number of patients who received at least one dose of asciminib. Percentages may not total 100 because of rounding. CML denotes chronic myeloid leukemia, and TKI tyrosine kinase inhibitor.

† Eastern Cooperative Oncology Group (ECOG) performance-status scores range from 0 to 5, with higher scores reflecting greater disability.

10 mg (1 patient), 20 mg (14 patients), 40 mg (35 patients), 80 mg (12 patients), 150 mg (13 patients), 160 mg (7 patients), and 200 mg (16 patients). Five dose-limiting toxic effects were reported: grade 3 elevations in the lipase level without clinical pancreatitis in 2 patients receiving 40 mg, grade 2 myalgia and arthralgia in 1 patient receiving 80 mg, grade 3 acute coronary syndrome in 1 patient receiving 150 mg, and grade 3 bronchospasm in 1 patient receiving 200 mg. In patients treated on a once-daily schedule, three dose levels were investigated: 80 mg (18 patients), 120 mg (22 patients), and 200 mg (12 patients). Three dose-limiting toxic effects occurred in patients receiving 200 mg: a grade 3 elevation in the lipase level associated with clinical pancreatitis, a grade 3 asymptomatic elevation in the lipase level, and grade 3 abdominal pain of undetermined cause.

Among all 150 patients who could be evaluated for safety, the most common nonhematologic adverse events that emerged during treatment regardless of dose schedule were asymptomatic elevations in the lipase or amylase level, rash, and constitutional symptoms (e.g., fatigue, nausea, headache, and arthralgia), of which 92% were grade 1 or 2. Hypertension, reported in 19% of patients, was the most commonly reported cardiovascular adverse event (Table 2).

Clinical pancreatitis, marked by abdominal pain and elevation in the lipase level and confirmed by abdominal imaging, occurred in 5 patients (3 patients receiving 80 mg twice daily, 1 receiving 150 mg twice daily, and 1 receiving 200 mg once daily); three cases were reported as serious adverse events (Table S3). Four of 5 patients had a single episode of pancreatitis, and 1 patient had two episodes of pancreatitis while receiving a reduced dose of asciminib. All cases resolved within 5 to 10 days after discontinuation of asciminib, and the sole patient rechallenged with asciminib was able to continue treatment at a lower dose. Three of 5 patients had had pancreatitis when using a previous TKI. Asymptomatic biochemical elevations in the lipase or amylase level occurred in 35 additional patients across all doses except 10 mg twice daily. These events were self-limited and did not progress to clinical pancreatitis. A total of 10 patients required temporary dose interruptions, and 1 patient discontinued treatment. Hematologic toxic effects that emerged during treatment

were common but were typically of grade 1 or 2 (Table 2).

PHARMACOKINETICS

For both twice-daily and once-daily schedules, the relationships between asciminib dose and both peak blood concentration and area under the curve for each dose level on days 1 and 29 were approximately dose proportional (Fig. S4A and S4B). The preliminary half-life was approximately 8 hours, which suggests a steady state by day 3. At a dose of 40 mg twice daily or 80 mg once daily, trough blood concentrations surpassed the preclinical 90% inhibitory concentration for phosphorylated signal transducer and activator of transcription 5 (pSTAT5) inhibition in a KCL-22 xenograft animal model (121 ng per milliliter). Pharmacokinetic–pharmacodynamic model–based predictions indicated that a dose of 40 mg twice daily would maintain 100% of patients without a T315I mutation above the preclinical 90% inhibitory concentration for pSTAT5 inhibition.

ANTILEUKEMIC ACTIVITY

Patients with Chronic-Phase CML without a T315I Mutation

Among 113 patients with chronic-phase CML without a T315I mutation receiving asciminib either once or twice daily, 34 of 37 patients (92%) without a complete hematologic response at baseline had a complete hematologic response (Table 3). Of 57 patients without a complete cytogenetic response at baseline, 31 (54%) had a complete cytogenetic response (Table 3) in a median time of 24 weeks (range, 4 to 126).

A major molecular response ($BCR-ABL1^{IS} \leq 0.1\%$) was achieved or maintained by 6 months in 37 of 99 patients (37%) who could be evaluated and by 12 months in 44 of 91 patients (48%) who could be evaluated (Table 3 and Table S4). Among the latter group of 91 patients, 30 of 40 patients (75%) with a baseline $BCR-ABL1^{IS}$ of 1% or less had a major molecular response by 12 months, whereas 14 of 51 patients (27%) with a $BCR-ABL1^{IS}$ of more than 1% had a major molecular response by 12 months. Of 85 patients without a deep molecular response (i.e., $BCR-ABL1^{IS} \leq 0.01\%$) at baseline, 17 (20%) had a deep molecular response by 12 months. By 12 months, 57 of 91 patients (63%) had an improvement in their molecular-response category (Table 4). In addition, a major molecular response was achieved

Table 2. Most Frequent Adverse Events That Emerged during Asciminib Monotherapy in Patients with Chronic-Phase or Accelerated-Phase CML.

Event	All Grades (N=150)	Grade 3 or 4 (N=150)
	number (percent)	
Total*	150 (100)	90 (60.0)
Fatigue	44 (29.3)	2 (1.3)
Headache	42 (28.0)	1 (0.7)
Lipase increased	40 (26.7)	15 (10.0)
Arthralgia	36 (24.0)	2 (1.3)
Nausea	36 (24.0)	1 (0.7)
Diarrhea	35 (23.3)	0
Rash	35 (23.3)	0
Thrombocytopenia	33 (22.0)	14 (9.3)
Vomiting	31 (20.7)	4 (2.7)
Hypertension	29 (19.3)	14 (9.3)
Upper respiratory tract infection	27 (18.0)	0
Abdominal pain	25 (16.7)	0
Pain in arm or leg	24 (16.0)	0
Pruritus	24 (16.0)	1 (0.7)
Back pain	23 (15.3)	2 (1.3)
Constipation	21 (14.0)	0
Pyrexia	21 (14.0)	0
Dizziness	20 (13.3)	1 (0.7)
Amylase increased	19 (12.7)	4 (2.7)
Cough	19 (12.7)	0
Dyspnea	19 (12.7)	2 (1.3)
Myalgia	19 (12.7)	1 (0.7)
Anemia	17 (11.3)	11 (7.3)
Hypertriglyceridemia	17 (11.3)	4 (2.7)
Nasopharyngitis	17 (11.3)	0
Alanine aminotransferase increased	16 (10.7)	4 (2.7)
Neutropenia	16 (10.7)	11 (7.3)
Abdominal pain, upper	15 (10.0)	0
Aspartate aminotransferase increased	15 (10.0)	3 (2.0)
Bone pain	15 (10.0)	1 (0.7)
Insomnia	15 (10.0)	1 (0.7)
Edema, peripheral	15 (10.0)	0
Hyperhidrosis	14 (9.3)	0
Hypophosphatemia	14 (9.3)	2 (1.3)
Hyperglycemia	13 (8.7)	3 (2.0)
Noncardiac chest pain	13 (8.7)	1 (0.7)
Decreased appetite	12 (8.0)	1 (0.7)
Depression	12 (8.0)	0
Dry eye	12 (8.0)	0

Table 2. (Continued.)

Event	All Grades (N = 150)	Grade 3 or 4 (N = 150)
	number (percent)	
γ-Glutamyltransferase increased	12 (8.0)	3 (2.0)
Hyperuricemia	12 (8.0)	2 (1.3)
Musculoskeletal pain	12 (8.0)	0
Vision blurred	12 (8.0)	0
Anxiety	11 (7.3)	2 (1.3)
Dry skin	11 (7.3)	0
Flank pain	11 (7.3)	0
Muscle spasms	11 (7.3)	0
Oropharyngeal pain	11 (7.3)	0
Weight increased	11 (7.3)	0
Fall	10 (6.7)	1 (0.7)
Dyspepsia	9 (6.0)	0
Hypokalemia	9 (6.0)	1 (0.7)
Influenza	9 (6.0)	1 (0.7)
Memory impairment	9 (6.0)	0
Pleural effusion	9 (6.0)	4 (2.7)
Abdominal discomfort	8 (5.3)	0
Blood creatinine increased	8 (5.3)	0
Urinary tract infection	8 (5.3)	1 (0.7)

* Data are for all patients who received at least one dose of asciminib.

or maintained by 12 months in 8 of 14 patients (57%) with chronic-phase CML who were deemed to have resistance to or unacceptable side effects from ponatinib.

Among 44 patients in whom a major molecular response was either achieved or maintained, all but 4 continued to receive treatment at the time of analysis; the median time in which a major molecular response was achieved was 20 weeks (range, 2 to 120), and the median duration of response was more than 61 weeks (range, 4 to 154). In these 4 patients, the major molecular response was lost between 28 and 100 weeks of treatment; 2 of them remained in the study with a complete cytogenetic response. A total of 21 of 64 patients (33%) without detectable mutations and 5 of 8 (62%) with mutations other than T315I had a major molecular response by 12 months. Hematologic, cytogenetic, and molecular responses were noted across all doses of asciminib administered on once-daily or twice-daily schedules.

Patients with Chronic-Phase CML with a T315I Mutation

Among 28 patients with chronic-phase CML harboring a T315I mutation, 14 of 16 (88%) without a complete hematologic response at baseline had a complete hematologic response (Table 3). Of 22 patients without a complete cytogenetic response at baseline, 9 (41%) had a complete cytogenetic response (Table 3) in a median time of 8 weeks (range, 4 to 33).

A major molecular response was achieved in 4 of 17 patients (24%) and maintained in 1 of 1 patient (100%) by 12 months, with 9 of 18 (50%) showing improvement in their molecular-response category by 12 months (Table 4). Of 5 patients with a T315I mutation who were deemed to have resistance to ponatinib, 1 (20%) had a major molecular response by 12 months. Three of 4 patients (75%) with chronic-phase CML with a T315I mutation who had a major molecular response received a dose of more than 150 mg twice daily (Table S5). All 5 patients

Table 3. Hematologic, Cytogenetic, and Molecular Responses with Asciminib (Combined Once-Daily and Twice-Daily Schedules).*

Variable	Chronic-Phase CML				Accelerated-Phase CML			
	No T3151 Mutation		T3151 Mutation		No T3151 Mutation		T3151 Mutation	
	Overall Response (N=113) †	Response Achieved (N=28) †	Response Maintained (N=15-167)	Response Achieved (N=28) †	Response Maintained (N=15-72)	Overall Response (N=4) †	Response Achieved (N=5) †	Response Maintained (N=5) †
Median follow-up (range) — wk	72 (0.1–167)	37 (0.7–167)		46 (15–72)	16 (6–120)			
Patients remaining in the study — no. (%)	88 (78)	19 (68)		2 (50)	1 (20)			
Complete hematologic response — no./total no. (%) ‡	34/37 (92)	14/16 (88)		3/3 (100)	4/5 (80)			
Major cytogenetic response — no./total no. (%) ‡§	85/110 (77)	61/70 (87)	15/25 (60)	11/20 (55)	4/5 (80)	0/4	1/5 (20)	0/1
Complete cytogenetic response — no./total no. (%) ‡§	77/110 (70)	46/53 (87)	11/25 (44)	9/22 (41)	2/3 (67)	0/4	1/5 (20)	0/1
Major molecular response — no./total no. (%) ‡¶								
In all patients								
By 6 mo	37/99 (37)	19/80 (24)	5/20 (25)	4/19 (21)	1/1 (100)	0/4	1/5 (20)	0
By 12 mo	44/91 (48)	26/72 (36)	5/18 (28)	4/17 (24)	1/1 (100)	0/4	1/5 (20)	0
In patients with ≤2 previous TKIs								
By 6 mo	13/25 (52)	5/15 (33)	4/10 (40)	3/9 (33)	1/1 (100)	0/1	1/5 (20)	0
By 12 mo	15/25 (60)	7/15 (47)	4/9 (44)	3/8 (38)	1/1 (100)	0/1	1/5 (20)	0
In patients with >2 previous TKIs**								
By 6 mo	24/74 (32)	14/64 (22)	1/10 (10)	1/10 (10)	0	0/3	1/5 (20)	0
By 12 mo	29/66 (44)	19/56 (34)	1/9 (11)	1/9 (11)	0	0/3	1/5 (20)	0

In patients with resistance to or unacceptable side effects from ponatinib††	7/17 (41)	3/13 (23)	4/4 (100)	4/4 (100)	1/7 (14)	1/7 (14)	0/0	0/2	0/2
By 6 mo									
By 12 mo	8/14 (57)	4/10 (40)	4/4 (100)	4/4 (100)	1/6 (17)	1/6 (17)	0/0	0/2	0/2

* For definitions of hematologic, cytogenetic, and molecular responses, see the Methods section in the Supplementary Appendix.

† Shown is the number of patients who received at least one dose of asciminib.

‡ The total number is the number of patients who could be evaluated.

§ Data on cytogenetic responses are based on patients who presented with Philadelphia chromosome-positive CML at baseline. Calculation of the number of patients in whom a major cytogenetic response or complete cytogenetic response was achieved is based on patients not in the respective response category at baseline.

¶ Molecular-response assessment is reported only for patients with the b2a2 or b3a2 transcripts; 7 patients had atypical BCR-ABL1 transcripts and were not included in the response assessment. The numbers of patients who received at least one dose of asciminib were as follows: 34 with chronic-phase CML without a T315I mutation, 12 with chronic-phase CML with a T315I mutation, and 1 with accelerated-phase CML without a T315I mutation.

** The numbers of patients who received at least one dose of asciminib were as follows: 79 with chronic-phase CML without a T315I mutation, 16 with chronic-phase CML with a T315I mutation, 3 with accelerated-phase CML without a T315I mutation, and 5 with accelerated-phase CML with a T315I mutation.

†† The numbers of patients who received at least one dose of asciminib were as follows: 18 with chronic-phase CML without a T315I mutation, 11 with chronic-phase CML with a T315I mutation, and 2 with accelerated-phase CML with a T315I mutation.

with a T315I mutation in whom a major molecular response was either achieved or maintained continued to receive treatment and were having a response at the time of analysis; the median time in which a major molecular response was achieved was 14 weeks (range, 4 to 20), and the median duration of response was more than 25 weeks (range, 12 to 96).

Patients with Accelerated-Phase CML

Among nine patients with accelerated-phase CML, seven of eight (88%) with hematologic disease at baseline had a complete hematologic response, and one of nine (11%) had a major molecular response, with responses maintained during therapy for a median of more than 11 weeks (Table 3).

Development of Myristoyl-Pocket Mutations

New myristoyl-pocket mutations were detected in 2 of 20 patients who had disease progression during asciminib treatment and in 2 of 66 patients without evidence of disease progression who had received asciminib for at least 12 months at the time of analysis. Four patients whose disease progressed before 12 months were not screened for mutations owing to sample unavailability. One patient with chronic-phase CML with a BCR-ABL1^{IS} of 8.1% and a baseline E255K mutation received asciminib at a dose of 40 mg twice daily. The patient had a major molecular response by 6 months but eventually discontinued the study with progressive disease associated with a new myristoyl-pocket G463S mutation at week 50 of treatment (Fig. S5). Details of the other 3 patients are presented in the Supplementary Appendix.

DISCUSSION

Asciminib had substantial and durable clinical activity in a heavily pretreated population of patients with chronic-phase or accelerated-phase CML in whom treatment with currently available ATP-competitive TKIs had failed. Major side effects were asymptomatic elevations in the lipase or amylase level, rash, and constitutional symptoms (e.g., fatigue, nausea, headache, and arthralgia), most of which were of grade 1 or 2 and appeared to be equivalent in patients receiving asciminib twice daily and in those receiving the drug once daily. A maximum tolerated dose was not reached. Clinical pancreatitis was noted in

Table 4. Categorical Response Shift from Baseline in Patients with Chronic-Phase CML Treated with Asciminib.*

Variable	No T3151 Mutation					T3151 Mutation				
	Baseline BCR-ABL1 ^{IS} †					Baseline BCR-ABL1 ^{IS} †				
	≤0.01% (N=6)	>0.01 to 0.1% (N=13)	>0.1 to 1% (N=22)	>1 to 10% (N=21)	>10% (N=42)	>0.01 to 0.1% (N=1)	>0.1 to 1% (N=2)	>1 to 10% (N=5)	>10% (N=19)	
Post-treatment BCR-ABL1^{IS} by 6 mo										
Patients who could be evaluated‡	6	13	23	18	39	1	1	5	13	
Distribution — no. of patients (%)§										
≤0.01%	6 (100)	4 (31)	5 (22)	4 (22)	1 (3)	1 (100)	0	0	0	
>0.01 to 0.1%	0	8 (62)	6 (26)	1 (6)	2 (5)	0	1 (100)	2 (40)	1 (8)	
>0.1 to 1%	0	1 (8)	12 (52)	12 (67)	7 (18)	0	0	2 (40)	1 (8)	
>1 to 10%	0	0	0	1 (6)	12 (31)	0	0	1 (20)	2 (15)	
>10%	0	0	0	0	17 (44)	0	0	0	9 (69)	
Post-treatment BCR-ABL1^{IS} by 12 mo										
Patients who could be evaluated‡	6	13	21	17	34	1	1	5	11	
Distribution — no. of patients (%)§										
≤0.01%	6 (100)	5 (38)	6 (29)	5 (29)	1 (3)	1 (100)	0	2 (40)	0	
>0.01 to 0.1%	0	7 (54)	6 (29)	3 (18)	5 (15)	0	1 (100)	0	1 (9)	
>0.1 to 1%	0	1 (8)	9 (43)	8 (47)	6 (18)	0	0	2 (40)	1 (9)	
>1 to 10%	0	0	0	1 (6)	12 (35)	0	0	1 (20)	1 (9)	
>10%	0	0	0	0	10 (29)	0	0	0	8 (73)	

* Percentages may not total 100 because of rounding. BCR-ABL1^{IS} denotes the ratio of BCR-ABL1 to ABL1 measured on the International Scale.

† The number of patients is the number who received at least one dose of asciminib in each category of BCR-ABL1 transcript level at baseline. A total of 10 patients with chronic-phase CML (9 without a T3151 mutation and 1 with a T3151 mutation) had a missing BCR-ABL1^{IS} value at baseline.

‡ Shown is the number of patients who could be evaluated in each category of baseline BCR-ABL1 transcript level who had undergone assessment of molecular response at 6 (or 12) months after treatment or who had a major molecular response within 6 (or 12) months. For a detailed definition of molecular response, see the Methods section in the Supplementary Appendix. Response assessment is reported only for patients with the b2a2 or b3a2 transcripts.

§ Percentages were calculated on the basis of the number of patients who could be evaluated.

3% of patients overall, only at asciminib doses of more than 40 mg twice daily, and was manageable with dose modifications. Myelosuppression was uncommon and mostly of grade 1 and 2. Among patients with chronic-phase CML without a T315I mutation, the incidences of complete cytogenetic response and major molecular response at 12 months were 70% and 48%, respectively. Among patients who entered the study with a *BCR-ABL1*¹⁵ of 0.1% or less at baseline, a deep molecular response was achieved or maintained in 60% during the study.

Although TKI therapy has transformed the natural history of CML,²⁵ many patients have TKI failure — frequently due to the emergence of resistance mutations in the *BCR-ABL1* kinase domain.^{8,9,26-29} In our study, asciminib showed activity in patients with or without *BCR-ABL1* kinase domain mutations.

Complete cytogenetic and major molecular responses were achieved in patients with chronic-phase CML with a T315I mutation, with the majority of those who had a response receiving asciminib doses of more than 150 mg twice daily, which was higher than the doses required to achieve responses in patients without a T315I mutation. This finding mirrors preclinical *in vitro* observations, in which the concentrations of asciminib that were required to achieve half the maximum inhibitory concentration were 5 to 10 times higher in cell lines expressing T315I-mutated *BCR-ABL1* than in cell lines expressing non-T315I-mutated *BCR-ABL1*.¹⁵ These clinical responses are important because, currently, only ponatinib yields meaningful clinical benefit for patients with a T315I mutation; however, the vas-

cular events that are associated with ponatinib often limit its use. Furthermore, a major molecular response was achieved in some patients with CML who were deemed to have resistance to or unacceptable side effects from ponatinib, which indicates a benefit in patients with limited effective therapeutic options other than stem-cell transplantation.

Asciminib was developed to bind to the myristoyl pocket — a previously unexploited feature of *ABL1* and *ABL2* kinases that is key to physiological autoinhibition of the native kinase. Given the extensive homology between the ATP-binding sites of many human kinases, targeting the myristoyl pocket is predicted to achieve superior selectivity and, hence, reduced toxicity. Asciminib exhibits both *in vitro* potency similar to that of second-generation TKIs and a resistance profile distinct from that of catalytic-site inhibitors.^{15,16} Although recent *in vitro* work suggested the potential for a high rate of emergent mutations with inhibition of the myristoyl pocket of *BCR-ABL1*,³⁰ our early clinical experience does not support this prediction. To date, myristoyl-pocket mutations have been detected in only 4 of 86 patients receiving asciminib, and clinical responses were maintained in the majority of patients who had a response.

In our study, asciminib monotherapy showed durable clinical activity in most patients with chronic-phase CML. Low-grade, reversible toxic effects occurred in a minority of patients.

A data sharing statement provided by the authors is available with the full text of this article at NEJM.org.

Supported by Novartis Pharmaceuticals.

Disclosure forms provided by the authors are available with the full text of this article at NEJM.org.

APPENDIX

The authors' full names and academic degrees are as follows: Timothy P. Hughes, M.D., Michael J. Mauro, M.D., Jorge E. Cortes, M.D., Hironobu Minami, M.D., Delphine Rea, M.D., Daniel J. DeAngelo, M.D., Ph.D., Massimo Breccia, M.D., Yeow-Tee Goh, M.D., Moshe Talpaz, M.D., Andreas Hochhaus, M.D., Philipp le Coutre, M.D., Oliver Ottmann, M.D., Michael C. Heinrich, M.D., Juan L. Steegmann, M.D., Ph.D., Michael W.N. Deininger, M.D., Ph.D., Jeroen J.W.M. Janssen, M.D., Ph.D., Francois-Xavier Mahon, M.D., Yosuke Minami, M.D., Ph.D., David Yeung, M.D., David M. Ross, M.B., B.S., Ph.D., Martin S. Tallman, M.D., Jae H. Park, M.D., Brian J. Druker, M.D., David Hynds, M.S., Yuyan Duan, Ph.D., Christophe Meille, Ph.D., Florence Hourcade-Potelleret, Ph.D., K. Gary Vanasse, M.D., Fabian Lang, M.D., and Dong-Wook Kim, M.D., Ph.D.

The authors' affiliations are as follows: the South Australian Health and Medical Research Institute and the University of Adelaide, Adelaide, SA, Australia (T.P.H., D.Y., D.M.R.); Memorial Sloan Kettering Cancer Center, New York (M.J.M., M.S.T., J.H.P.); University of Texas M.D. Anderson Cancer Center, Houston (J.E.C.); Kobe University Graduate School of Medicine, Kobe (H.M.), and the National Cancer Center Hospital East, Chiba (Y.M.) — both in Japan; Hôpital Saint-Louis, Paris (D.R.), and the University of Bordeaux, Bordeaux (F.-X.M.) — both in France; Dana-Farber Cancer Institute, Boston (D.J.D.); Sapienza University, Rome (M.B.); Singapore General Hospital, Singapore (Y.-T.G.); University of Michigan Comprehensive Cancer Center, Ann Arbor (M.T.); Universitätsklinikum Jena, Jena (A.H.), Charité Hospital, Berlin (P.C.), and the Department for Hematology-Oncology, Goethe University Hospital, Frankfurt am Main (F.L.) — all in Germany; University of Cardiff, Cardiff, United Kingdom (O.O.); Veterans Affairs Portland Health Care System (M.C.H.) and Oregon Health and Science University Knight Cancer Institute (M.C.H., B.J.D.), Portland; Hospital de la Princesa and Instituto de Investigación Sanitaria Princesa, Madrid (J.L.S.); Huntsman Cancer Institute, University of Utah, Salt Lake City (M.W.N.D.); Amsterdam University Medical Centers, VU University Medical Center, Amsterdam (J.J.W.M.J.); Novartis Pharma, Basel, Switzerland (D.H., Y.D., C.M., F.H.-P., K.G.V.); and Seoul St. Mary's Hematology Hospital, Catholic University of Korea, Seoul, South Korea (D.-W.K.).

REFERENCES

- Hughes TP, Saglio G, Quintás-Cardama A, et al. BCR-ABL1 mutation development during first-line treatment with dasatinib or imatinib for chronic myeloid leukemia in chronic phase. *Leukemia* 2015;29:1832-8.
- Shah NP, Nicoll JM, Nagar B, et al. Multiple BCR-ABL kinase domain mutations confer polyclonal resistance to the tyrosine kinase inhibitor imatinib (STI571) in chronic phase and blast crisis chronic myeloid leukemia. *Cancer Cell* 2002;2:117-25.
- Branford S, Rudzki Z, Walsh S, et al. Detection of BCR-ABL mutations in patients with CML treated with imatinib is virtually always accompanied by clinical resistance, and mutations in the ATP phosphate-binding loop (P-loop) are associated with a poor prognosis. *Blood* 2003;102:276-83.
- Kantarjian HM, Shah NP, Cortes JE, et al. Dasatinib or imatinib in newly diagnosed chronic-phase chronic myeloid leukemia: 2-year follow-up from a randomized phase 3 trial (DASISION). *Blood* 2012;119:1123-9.
- Cortes J, Jabbour E, Kantarjian H, et al. Dynamics of BCR-ABL kinase domain mutations in chronic myeloid leukemia after sequential treatment with multiple tyrosine kinase inhibitors. *Blood* 2007;110:4005-11.
- Kantarjian HM, Hochhaus A, Saglio G, et al. Nilotinib versus imatinib for the treatment of patients with newly diagnosed chronic phase, Philadelphia chromosome-positive, chronic myeloid leukaemia: 24-month minimum follow-up of the phase 3 randomised ENESTnd trial. *Lancet Oncol* 2011;12:841-51.
- Carter TA, Wodicka LM, Shah NP, et al. Inhibition of drug-resistant mutants of ABL, KIT, and EGF receptor kinases. *Proc Natl Acad Sci U S A* 2005;102:11011-6.
- O'Hare T, Eide CA, Deininger MW. Bcr-Abl kinase domain mutations, drug resistance, and the road to a cure for chronic myeloid leukemia. *Blood* 2007;110:2242-9.
- O'Hare T, Shakespeare WC, Zhu X, et al. AP24534, a pan-BCR-ABL inhibitor for chronic myeloid leukemia, potently inhibits the T315I mutant and overcomes mutation-based resistance. *Cancer Cell* 2009;16:401-12.
- Valent P, Hadzijušević E, Scherthanner GH, Wolf D, Rea D, le Coutre P. Vascular safety issues in CML patients treated with BCR/ABL1 kinase inhibitors. *Blood* 2015;125:901-6.
- Moslehi JJ, Deininger M. Tyrosine kinase inhibitor-associated cardiovascular toxicity in chronic myeloid leukemia. *J Clin Oncol* 2015;33:4210-8.
- Saussele S, Krauss MP, Hehlmann R, et al. Impact of comorbidities on overall survival in patients with chronic myeloid leukemia: results of the randomized CML study IV. *Blood* 2015;126:42-9.
- Caldemeyer L, Dugan M, Edwards J, Akard L. Long-term side effects of tyrosine kinase inhibitors in chronic myeloid leukemia. *Curr Hematol Malig Rep* 2016;11:71-9.
- Stegmann JL, Baccarani M, Breccia M, et al. European LeukemiaNet recommendations for the management and avoidance of adverse events of treatment in chronic myeloid leukaemia. *Leukemia* 2016;30:1648-71.
- Wylie AA, Schoepfer J, Jahnke W, et al. The allosteric inhibitor ABL001 enables dual targeting of BCR-ABL1. *Nature* 2017;543:733-7.
- Schoepfer J, Jahnke W, Berellini G, et al. Discovery of asciminib (ABL001), an allosteric inhibitor of the tyrosine kinase activity of BCR-ABL1. *J Med Chem* 2018;61:8120-35.
- Baccarani M, Cortes J, Pane F, et al. Chronic myeloid leukemia: an update of concepts and management recommendations of European LeukemiaNet. *J Clin Oncol* 2009;27:6041-51.
- Talpaz M, Shah NP, Kantarjian H, et al. Dasatinib in imatinib-resistant Philadelphia chromosome-positive leukemias. *N Engl J Med* 2006;354:2531-41.
- Branford S, Fletcher L, Cross NC, et al. Desirable performance characteristics for BCR-ABL measurement on an international reporting scale to allow consistent interpretation of individual patient response and comparison of response rates between clinical trials. *Blood* 2008;112:3330-8.
- Kantarjian H, Sawyers C, Hochhaus A, et al. Hematologic and cytogenetic responses to imatinib mesylate in chronic myelogenous leukemia. *N Engl J Med* 2002;346:645-52.
- Cheson BD, Bennett JM, Kopecky KJ, et al. Revised recommendations of the International Working Group for Diagnosis, Standardization of Response Criteria, Treatment Outcomes, and Reporting Standards for Therapeutic Trials in Acute Myeloid Leukemia. *J Clin Oncol* 2003;21:4642-9.
- Hughes T, Deininger M, Hochhaus A, et al. Monitoring CML patients responding to treatment with tyrosine kinase inhibitors: review and recommendations for harmonizing current methodology for detecting BCR-ABL transcripts and kinase domain mutations and for expressing results. *Blood* 2006;108:28-37.
- Babb J, Rogatko A, Zacks S. Cancer phase I clinical trials: efficient dose escalation with overdose control. *Stat Med* 1998;17:1103-20.
- Neuenschwander B, Branson M, Gsponer T. Critical aspects of the Bayesian approach to phase I cancer trials. *Stat Med* 2008;27:2420-39.
- Kantarjian H, O'Brien S, Jabbour E, et al. Improved survival in chronic myeloid leukemia since the introduction of imatinib therapy: a single-institution historical experience. *Blood* 2012;119:1981-7.
- Cortes JE, Kantarjian H, Shah NP, et al. Ponatinib in refractory Philadelphia chromosome-positive leukemias. *N Engl J Med* 2012;367:2075-88.
- Quintás-Cardama A, Kantarjian HM, Cortes JE. Mechanisms of primary and secondary resistance to imatinib in chronic myeloid leukemia. *Cancer Control* 2009;16:122-31.
- Apsel Winger B, Shah NP. PPAR γ : welcoming the new kid on the CML stem cell block. *Cancer Cell* 2015;28:409-11.
- Khorashad JS, Anand M, Marin D, et al. The presence of a BCR-ABL mutant allele in CML does not always explain clinical resistance to imatinib. *Leukemia* 2006;20:658-63.
- Lee BJ, Shah NP. Identification and characterization of activating ABL1 1b kinase mutations: impact on sensitivity to ATP-competitive and allosteric ABL1 inhibitors. *Leukemia* 2017;31:1096-107.

Copyright © 2019 Massachusetts Medical Society.